

Manual DNA Extraction from Food Samples

Manual DNA purification from food samples using the ReliaPrep™ Blood gDNA Miniprep System followed by amplification using GoTaq® qPCR Master Mix.

Kit:	ReliaPrep™ Blood gDNA Miniprep System (Cat.# A5081)
Analyses:	Quantitation by absorbance and with fluorescent dye; qPCR amplification
Sample Type(s):	Ground seed (corn and wheat) and meat samples (pork and beef)
Input:	50–100mg
Materials Required:	

- ReliaPrep™ Blood gDNA Miniprep System (Cat.# A5081)
- CTAB Buffer (Cat.# MC1411)
- RNase A Solution (Cat.# A7973)
- Proteinase K (PK) Solution (Part# A505C)
- heat block
- microcentrifuge
- 100% isopropanol
- Elution Buffer (Cat.# A8281)
- GoTaq® qPCR Master Mix (Cat.# A6002)
- QuantiFluor® ONE dsDNA System (Cat.# E4871)

This protocol was developed by Promega Applications Scientists and is intended for research use only.

Users are responsible for determining suitability of the protocol for their application.

For further information, e-mail technical services at: techserv@promega.com

Protocol:

1. For seed samples: Add 1ml of CTAB Buffer, 20µl of RNase A Solution and 40µl of Proteinase K (PK) Solution to each tube containing up to 100mg of sample. Vortex until completely suspended.
2. For meat samples: Add 600µl of CTAB Buffer, 2µl of RNase Solution and 30µl of Proteinase K (PK) Solution to each tube containing up to 100mg of sample. Vortex until completely suspended.
3. Place samples in a heat block at 65°C (seed samples) or 60°C (meat samples) for 30 minutes. After incubation, vortex to mix. Centrifuge samples for 10 minutes at $\geq 16,000 \times g$.
4. Transfer 300µl of clear supernatant to a clean 1.5ml microcentrifuge tube.
5. Add 300µl of CLD Buffer to the cleared supernatant and mix. Add 600µl of 100% isopropanol and vortex.
6. Load 600µl of sample onto a ReliaPrep™ Binding Column placed in a collection tube. Centrifuge for 1 minute at maximum speed. Discard flowthrough.
7. Load the rest of the sample to the ReliaPrep™ Binding Column and centrifuge for 1 minute. Place Binding Column into a new collection tube.
8. Add 500µl of Column Wash Solution (CWD). Centrifuge for 2 minutes at maximum speed. Discard the flowthrough.
9. Repeat Step 7 twice for a total of three washes.
10. Place Binding Column in a labeled elution tube. Add 100µl of Elution Buffer to the Binding Column. Centrifuge for 1 minute at maximum speed. Discard the column and save eluate.

Results:

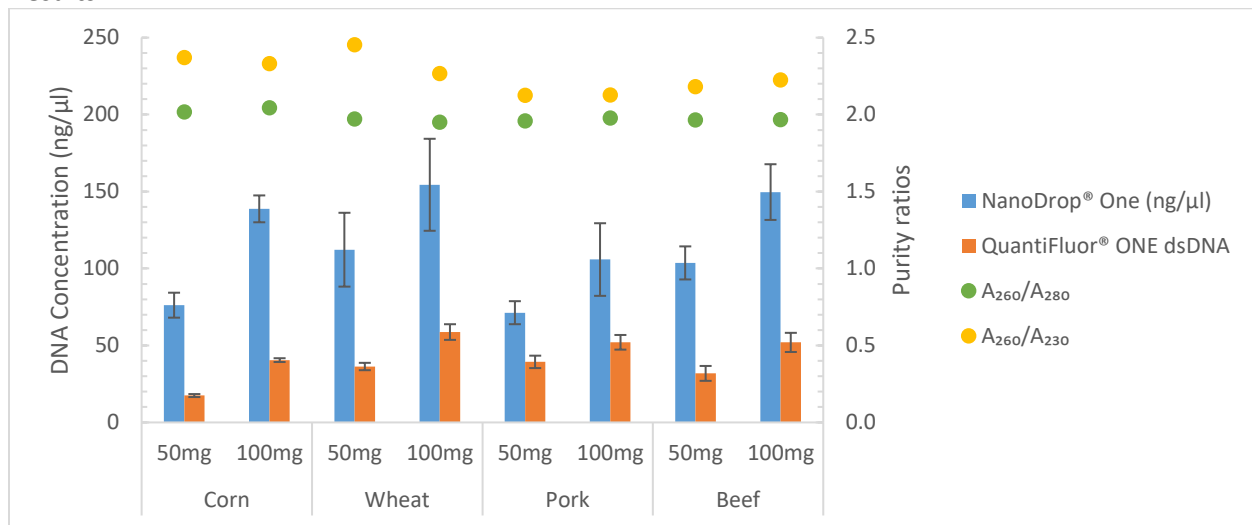


Figure 1. DNA concentration and purity ratios obtained for DNA extracted from 50mg or 100mg of food samples using the ReliaPrep™ Blood gDNA Miniprep System. DNA concentration and purity ratios were assessed by absorbance with NanoDrop® One Spectrophotometer and by using the QuantiFluor® ONE dsDNA System (Cat.# E4871). Error bars indicate standard deviation (N=3).

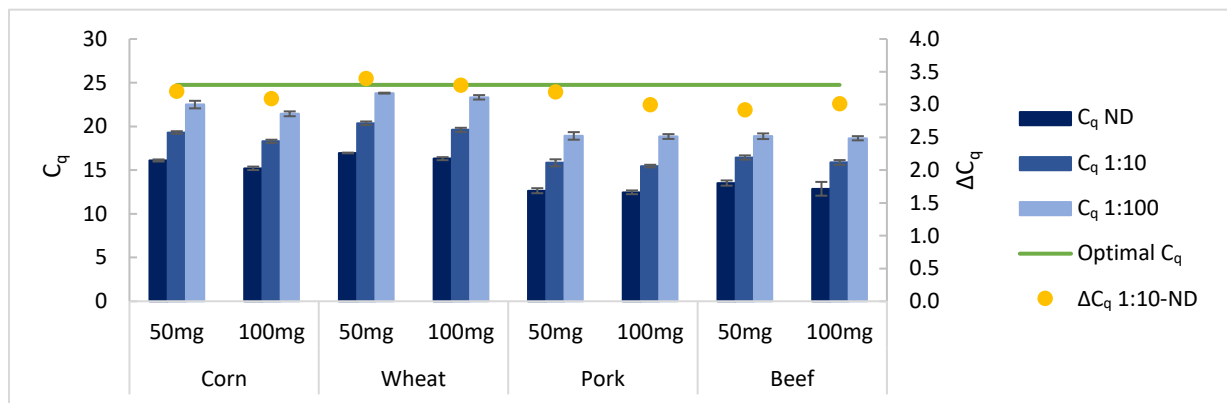


Figure 2. qPCR amplification results for DNA extracted from 50mg or 100mg of food samples using ReliaPrep™ Blood gDNA Miniprep System. C_q and ΔC_q values for 2μl of the eluted DNA amplified using GoTaq® qPCR Master Mix (Cat.# A6001) and universal plant primers (1) or pork and beef primers (2) in a final volume of 20μl (final concentration of primers 500nM). A ΔC_q value of 3.3 indicates no qPCR inhibitor compounds are present.

References:

1. Wang, J. *et al.* (2011) Universal endogenous gene controls for bisulphite conversion in analysis of plant DNA methylation. *Plant Methods* **7**, 39.
2. López-Andreo, M. *et al.* (2005) Identification and quantitation of species in complex DNA mixture by real-time polymerase chain reaction. *Anal. Biochem.* **339**, 73–82.